

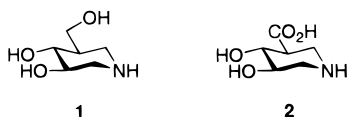
Highly Selective Synthesis of 1-*N*-Iminosugars of the D-Glucose and -Glucuronic Acid Types

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Members of a new class of iminosugars, the 1-*N*-iminosugars, have been known to act as specific and potent inhibitors of β -glycosidases.^{1–3} [In this text, we use a nomenclature convention in which compounds with a nitrogen atom at the anomeric position (piperidine derivatives) are named as “1-*N*-iminosugars”. They are also called isofagomine derivatives or 1-azasugars.] Although these 1-*N*-iminosugars have great potential as useful molecular probes to study the biological implication of β -glycosidases, their syntheses are rather tedious because of the lack of a well-established method for introducing the aminomethyl (or its equivalent) group required to produce the 1-*N*-iminosugar. In our ongoing program of developing inhibitors of glycozymes (enzymes involved in carbohydrate biosynthetic and catabolic pathways), we needed to establish a general synthetic route to provide these iminosugars in large quantity for further study of their biological activity. Herein we describe an efficient synthetic approach that provides the easy access to 1-*N*-iminosugar derivatives of glucose **1** and glucuronic acid **2**, starting from a readily obtainable (*R*)-2,3-*O*-cyclohexylidene-glyceraldehyde **3**.⁴



Synthesis of D-Glucose Type 1-*N*-Iminosugar 1. Horner-Emmons condensation⁵ of the (*R*)-2,3-*O*-cyclohexylidene-glyceraldehyde **3**⁴ using trimethylphosphonoacetate and sodium hydride afforded the α,β -unsaturated methyl ester **4** in an *E/Z* ratio of 10:1 in 83% yield⁶ (Scheme 1). Treatment of **4** with DIBAL in CH₂Cl₂ gave the allylic alcohol **5**, which was subjected to Sharpless

Table 1. Nucleophilic Epoxy Ring Opening Reaction of **6** with CN⁻

entry	conditions	yield	product ratio (7a:7b)
1	Et ₂ AlCN/toluene/rt/48 h	91%	1:1
2	LiCN/THF/reflux/5 h	50%	10:1
3	KCN/Bu ₄ Ni/Ti(O- <i>i</i> Pr) ₄ /DMSO/rt/60 h	>90%	12:1

asymmetric epoxidation⁷ with diethyl L-(+)-tartrate to afford the epoxy alcohol **6** in 90% yield.

Because our strategy was to introduce a cyanide group in a diastereo- and regioselective manner to construct the 1-*N*-iminosugar framework, we examined the regioselective ring opening reaction of the epoxide **6** with a cyanide anion (Table 1). When Nagata's reagent,⁸ diethylaluminum cyanide (Et₂AlCN), was applied, an excellent yield of a cyano diol derivative was obtained. However, almost no regioselectivity was observed, and the reaction afforded the desired 2-cyano derivative **7a** and 3-cyano derivative **7b** in a ratio of 1:1⁹ (entry 1). The regioselectivity was determined on the basis of the ¹H NMR spectrum of the product, in which the H-2 of the 2-CN isomer **7a** appeared at δ 2.93 ppm as a doublet of triplets, and the H-3 of the 3-CN isomer **7b** appeared at δ 3.13 ppm as a doublet of doublets (Figure 1). This result was consistent with our previous observation in the reaction of (2*R*,3*S*)-3-[(trityl)oxy]oxiranemethanol with Et₂AlCN, in which the C-3 attack was favorable.¹⁰

When the epoxide was treated with a highly reactive LiCN¹¹ prepared in situ from lithium hydride and acetone cyanohydrin, the desired product **7a** was obtained in good regioselectivity (10:1) but in low chemical yield as a result of a competing side reaction¹² (50% based on recovered starting material) (entry 2).

Good regioselectivity was obtained by treating the epoxide **6** under the Sharpless conditions,¹³ with KCN and Bu₄Ni in the presence of Ti(O-*i*Pr)₄ (entry 3). Although the reaction proceeded rather slowly (60 h at room temperature), an enhanced chemical yield and regioselectivity were observed, with **7a** and **7b** being formed in a ratio of 12:1 (>90% yield).

We speculated that the addition of Ti(O-*i*Pr)₄ to **6** could form a relatively rigid complex such as complex A (Figure 1),¹⁴ in which the backside of the C-3 position of the

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(9) Although **7a** and **7b** were separable on silica gel TLC (**7a**, *R*_f = 0.31 and **7b**, *R*_f = 0.21 in CHCl₃–EtOAc–MeOH 10:1:1), we purified the desired regioisomer (the 2-CN derivative) after the silylation as **8**. For the comparison, the two regioisomers **7a** and **7b** were differentiated by their ¹H NMR spectra in which the methine proton at the carbon attached to the CN appears at δ 2.93 ppm as a doublet of triplets (H-2) for **7a**, whereas for **7b** it appears at δ 3.13 ppm as a doublet of doublets (H-3).

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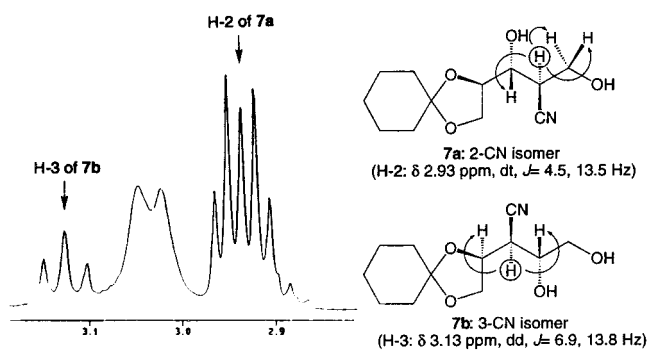
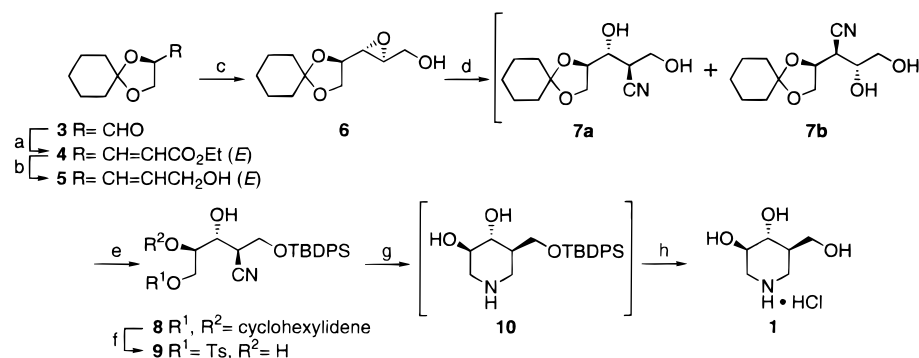
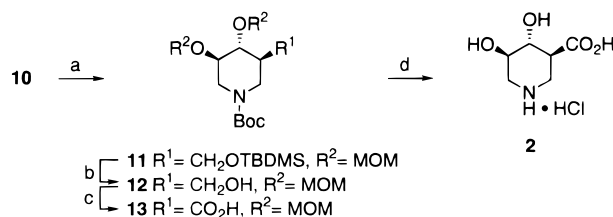
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(6) They were easily separable on silica gel chromatography. *R*_f values for the *E*- and *Z*-isomers were 0.5 and 0.6 in EtOAc–hexanes (1:4), respectively.

Scheme 1^aScheme 2^a

^a Reagents and conditions: (a) Boc₂O/Et₃N/MeOH (62% in two steps from **9**); (b) (1) MOMCl/*i*Pr₂NEt/CH₂Cl₂, (2) Bu₄NF/THF (74% in two steps); (c) (1) (COCl)₂/DMSO/Et₃N/CH₂Cl₂ (2) H₂O₂/NaClO₂/NaHPO₄/CH₃CN-H₂O (82% in two steps); (d) 3 N HCl (89%).

Synthesis of D-Glucuronic Acid Type 1-*N*-Iminosugar **2.** The D-glucuronic acid type 1-*N*-imosugar **2** was also prepared from the synthetic intermediate **10**, which was treated this time with Boc₂O to give **11** as a single product (Scheme 2). The diol moiety was protected with methoxymethyl (MOM) groups, and the silyl protecting group was removed with Bu₄NF to give **12**. Oxidation to the corresponding uronic acid derivative **13** was carried out stepwise:^{1b} first with DMSO and (COCl)₂,¹⁶ giving an aldehyde group, and further with H₂O₂ and NaClO₂¹⁷ in a CH₃CN-phosphate buffer to produce a D-glucuronic acid derivative **13**. All of the protecting groups of **13** were removed by 3 N HCl to provide the glucuronic acid type 1-*N*-imosugar **2**.^{1b}

In summary, we have described an efficient synthetic procedure for 1-*N*-imosugars of D-glucose **1** (25% overall yield) and D-glucuronic acid **2** (14% overall yield) from readily obtainable (*R*)-2,3-*O*-cyclohexylidene-glyceraldehyde employing Sharpless asymmetric epoxidation and the regioselective epoxide ring opening reaction with cyanide.

This synthetic route has allowed preparation of substantial amounts of 1-*N*-imosugars for further study of their effects in biological systems in which glycozymes play a key role.

Experimental Section

General Methods. The reagents used were purchased from Aldrich, Sigma, or Acros, and the solvents were reagent grade and used as supplied. ¹H NMR spectra were recorded at 300

Figure 1. ¹H NMR spectrum of a mixture of **7a** and **7b** used to determine the regioselectivity of the reaction of **6** and CN⁻ and a proposed conformation of the complex formed between **6** and Ti(O*i*Pr)₄.

epoxide **6** was blocked by a hydrogen of the C-5 methylene group; in contrast, the C-2 position would be well opened to the CN⁻ backside-attack by the complex formation of the hydroxymethyl and the epoxide oxygen groups with the titanium.

A mixture of **7a** and **7b** was treated with *tert*-butylchlorodiphenyl silane (TBDPSCI)¹⁵ to give **8** in high yield. The cyclohexylidene group in **8** was selectively removed with 80% acetic acid to afford the corresponding triol. The triol derivative was selectively tosylated at the C-5 primary alcohol with *p*-toluenesulfonyl chloride in pyridine to give **9** in good yield. Hydrogenation with Raney Ni first reduced the nitrile group to the corresponding aminomethyl group, which then displaced the 5-OTs group and allowed the intramolecular cyclization to proceed to give a silyl-protected 1-*N*-imosugar derivative **10**. Finally, treatment of **10** with 2 N HCl gave the D-glucose type 1-*N*-imosugar **1**.^{1b,2a}

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MHz, and ^{13}C NMR spectra were recorded at 75 MHz. Internal standards used in ^1H NMR spectra were TMS (δ 0.00) for CDCl_3 and HOD (δ 4.78) for D_2O and in ^{13}C NMR were CDCl_3 (δ 75.0) for CDCl_3 and CH_3CN (δ 1.30) for D_2O . Mass spectral data were analyzed by the Mass Spectrometry Laboratory at University of Illinois and Mass Spectrometry Facility at Johns Hopkins University.

Methyl (4S)-4,5-(Cyclohexylidenedioxy)-(2E)-pentenoate (4). To a mixture of NaH (6.5 g, 163.8 mmol) in benzene (80 mL) was added a solution of trimethylphosphonoacetate (26 mL, 180 mmol) in THF (100 mL) at 0°C , and the mixture was stirred at the same temperature for 1 h. Then, a solution of (*R*)-2,3-*O*-cyclohexylidene glyceraldehyde **3⁴** (14.0 g, 81.9 mmol) in THF (80 mL) was added, and the resulting mixture was allowed to warm to room temperature and stirred for 2 h. The reaction mixture was poured into ice water (250 mL) and extracted with EtOAc. The combined extracts were washed with brine, dried (MgSO_4), and concentrated. The residue was chromatographed on silica gel with EtOAc–hexane (1:6) to give the *E*-isomer **4** (14.1 g, 76%) and *Z*-isomer (1.4 g, 7%) as a colorless oil: ^1H NMR (CDCl_3) δ 1.35 (m, 3H, cyclohexylidene group), 1.56 (m, 7H, cyclohexylidene group), 3.61 (dd, 1H, $J = 7.5$ Hz), 3.69 (s, 3H), 4.12 (dd, 1H, $J = 6.9, 7.8$ Hz), 4.61 (dd, 1H, $J = 5.7, 3$ Hz), 6.06 (d, 1H, $J = 15.3$ Hz), 6.83 (dd, 1H, $J = 5.4, 15.3$ Hz); ^{13}C NMR (CDCl_3) δ 23.5, 23.6, 24.8, 35.0, 35.8, 51.3, 68.1, 74.3, 110.4, 121.4, 145.2, 166.0; HRMS FAB calcd for $\text{C}_{12}\text{H}_{18}\text{O}_4$ (M^+) 226.1205, found 226.1208.

(4S)-4,5-(Cyclohexylidenedioxy)-(2E)-penten-1-ol (5). To a solution of **4** (5.0 g, 22 mmol) in CH_2Cl_2 (150 mL) under argon was added a 1.0 M solution of DIBAL in toluene (44.0 mL, 44 mmol) at 0°C , and the mixture was stirred for 2 h at room temperature. Acetone (20 mL) was slowly added to the reaction mixture. An aqueous sodium potassium tartrate solution (100 mL) and EtOAc (200 mL) were subsequently added, and the resulting mixture was stirred until the layers were clearly separated. The aqueous layer was extracted with EtOAc, and the combined extracts were washed with brine, dried (MgSO_4), and concentrated. The residue was chromatographed on silica gel with EtOAc–hexane (1:3) to give **5** (4.0 g, 92%) as a colorless oil: ^1H NMR (CDCl_3) δ 1.24 (m, 3H, cyclohexylidene group), 1.46 (m, 7H, cyclohexylidene group), 3.42 (m, 2H, 1H of hydroxyl group), 3.93 (m, 3H), 4.37 (dd, 1H, $J = 7.2, 13.8$ Hz), 5.53 (dd, 1H, $J = 7.5, 15.6$ Hz), 5.76 (dt, 1H, $J = 5.1, 9.9, 15.6$ Hz); ^{13}C NMR (CDCl_3) δ 23.5, 23.5, 24.7, 35.0, 35.9, 61.7, 68.6, 75.8, 109.6, 127.9, 133.1; HRMS FAB calcd for $\text{C}_{11}\text{H}_{18}\text{O}_3$ (M^+) 198.1256, found 198.1259.

(2S,3R,4R)-2,3-Epoxy-4,5-(cyclohexylidenedioxy)pentan-1-ol (6). To a slurry of flame-dried powdered MS4A (15 g) in dry CH_2Cl_2 (400 mL) under argon were sequentially added titanium tetrakispropoxide (23.4 mL, 78.6 mmol) and diethyl *L*-(+)-tartrate (13.5 mL, 78.6 mmol) at -20°C , and the mixture was stirred for 30 min. A solution of **5** (10.4 g, 52.4 mmol) in dry CH_2Cl_2 (120 mL) was added, and the resulting mixture was stirred at -20°C for 30 min. A solution of *tert*-butylhydroperoxide in decane (5.0–6.0M, 21 mL, 104.8 mmol) was subsequently added dropwise to the mixture, and the resulting mixture was kept for 16 h at -15°C . Aqueous tartaric acid (10%, 200 mL) was added at -20°C , and the whole was allowed to warm to room temperature. After being stirred for 1 h, the reaction mixture was filtered, and the filtrate was extracted with CH_2Cl_2 . The combined extracts were washed with brine, dried (MgSO_4), and concentrated. The residue was chromatographed on silica gel with EtOAc–hexane (1:6) to give **6** (10.1 g, 90%) as a colorless oil: ^1H NMR (CDCl_3) δ 1.30 (br s, 3H, cyclohexylidene group), 1.51 (m, 7H, cyclohexylidene group), 2.01 (br s, 1H, hydroxyl group), 2.97 (dd, 1H, $J = 2.4, 4.8$ Hz), 3.03 (dd, 2H, $J = 2.4, 4.8$ Hz, 1H of hydroxyl group), 3.54 (dd, 1H, $J = 3.9, 12.6$ Hz), 3.74 (m, 1H), 3.81 (dd, 1H, $J = 1.8, 12.9$ Hz), 4.00 (m, 2H); ^{13}C NMR (CDCl_3) δ 23.5, 23.7, 24.8, 34.8, 35.7, 55.1, 55.4, 60.8, 65.4, 74.6, 110.3; HRMS FAB calcd for $\text{C}_{11}\text{H}_{18}\text{O}_4$ (M^+) 214.1205, found 214.1207.

(2R,3R,4R)-1-[(*tert*-Butyldiphenylsilyloxy)]-2-cyano-4,5-(cyclohexylidenedioxy)-3-hydroxypentane (8). To a stirred solution of **6** (3.3 g, 15.4 mmol) in DMSO (60 mL) under argon were added potassium cyanide (2.5 g, 38.5 mmol) and tetrabutylammonium iodide (10.2 g, 27.7 mmol) at room temperature, and then titanium tetrakispropoxide (11.4 mL, 38.5 mmol) was

slowly added. The reaction mixture was stirred at room temperature for 60 h, diluted with EtOAc (200 mL), and poured into aqueous NaHCO_3 solution (100 mL). The resulting mixture was extracted with EtOAc, and the combined extracts were washed with brine, dried (MgSO_4), and concentrated. This was employed for the next step without further purification.

To a solution of the above mixture (4.0 g) in DMF (60 mL) were added imidazole (3.4 g, 43.1 mmol) and *tert*-butylchlorodiphenyl silane (5.1 mL, 18.5 mmol) at $0-5^\circ\text{C}$, and the mixture was stirred for 2 h at room temperature. The reaction mixture was poured into ice-cooled aqueous NaHCO_3 solution (150 mL) and extracted with Et_2O . The combined extracts were successively washed with aqueous NaHCO_3 solution and brine, dried (MgSO_4), and concentrated. The residue was chromatographed on silica gel with EtOAc–hexane (1:30) to give the **8** (6.5 g, 88% in 2 steps) as a colorless oil: ^1H NMR (CDCl_3) δ 1.14 (s, 9H), 1.44 (m, 3H, cyclohexylidene group), 1.51 (m, 7H, cyclohexylidene group), 2.35 (d, 1H, $J = 8.7$ Hz), 2.88 (dt, 1H, $J = 4.5, 13.8$ Hz), 3.95 (m, 3H), 4.11 (m, 2H), 4.45 (dt, $J = 2.1, 6.3, 8.7$ Hz), 7.43 (m, 6H), 7.77 (m, 4H); ^{13}C NMR (CDCl_3) δ 19.3, 23.6, 23.9, 25.0, 26.7, 34.2, 35.9, 39.1, 60.6, 65.7, 68.1, 75.3, 110.5, 118.6, 27.8, 129.9, 123.0, 135.6, 135.6; HRMS FAB calcd for $\text{C}_{28}\text{H}_{38}\text{NO}_4\text{Si}$ ($M + \text{H}^+$) 480.2570, found 480.2571.

(2R,3R,4S)-1-[(*tert*-Butyldiphenylsilyloxy)]-2-cyano-3,4-dihydroxy-5-[(*p*-toluenesulfonyloxy)pentane (9). A solution of **8** (2.0 g, 4.2 mmol) in 80% acetic acid (30 mL) was stirred at room temperature for 40 h. The resulting reaction mixture was then concentrated and coevaporated with toluene (30 mL \times 2), and the residue was employed for the next step without further purification.

To a stirred solution of the above residue (1.7 g) in pyridine (40 mL) was added *p*-toluenesulfonyl chloride (1.0 g, 5.5 mmol) in one portion at room temperature, and the resulting mixture was stirred for 4 h at room temperature. The reaction mixture was poured into ice water (100 mL) and extracted with EtOAc. The combined extracts were successively washed with aqueous NaHCO_3 solution and brine, dried (MgSO_4), and concentrated. The residue was chromatographed on silica gel with EtOAc–hexane (1:4) to give **9** (1.6 g, 70% in 2 steps) as a colorless oil: ^1H NMR (CDCl_3) δ 1.01 (s, 9H), 2.44 (s, 3H), 3.00 (dt, 2H, $J = 4.5, 13.5$ Hz), 3.39 (br, 1H, hydroxyl group), 3.98 (m, 3H), 4.17 (bs, 3H), 7.34 (d, 2H, $J = 8.4$ Hz), 7.42 (m, 6H), 7.70 (m, 4H), 7.82 (d, 2H, $J = 8.4$ Hz); ^{13}C NMR (CDCl_3) δ 19.1, 21.5, 26.6, 37.5, 60.5, 67.9, 68.8, 70.3, 118.4, 127.6, 129.9, 132.2, 132.3, 135.4, 135.5, 145.2; HRMS FAB calcd for $\text{C}_{29}\text{H}_{36}\text{NO}_6\text{Si}$ ($M + \text{H}^+$) 554.2032, found 554.2031.

***D*-Glucose Type 1-*N*-Iminosugar: (3R,4R,5R)-5-(Hydroxymethyl)piperidine-3,4-diol Hydrochloride Salt (1).** A solution of **9** (2.0 g, 3.6 mmol) and Raney Ni (5 mL) in EtOH (20 mL) was stirred vigorously under H_2 atmosphere at room temperature for 50 h. After removal of the catalyst by filtration, the filtrate was concentrated to give a crude product **10**, which was employed for the next step without further purification.

A solution of the residue **10** in 2 N HCl (15 mL) was stirred overnight at room temperature, and the reaction mixture was concentrated. The residue was diluted with water (10 mL) and applied onto a column of Dowex 50W-X8 [H^+] resin. The column was washed with water, and the product was eluted out with 3% NH_4OH . The fractions containing **1** were pooled and concentrated. The residue was chromatographed on silica gel with 2-propanol– H_2O – NH_4OH (7:2:1) to give chromatographically pure **1**. Water (10 mL) and 1 N HCl (20 mL) were added to the residue, and the solution was concentrated to form a hydrochloride salt of the iminosugar. The residue was applied onto a column of Sephadex G-25 (2 cm \times 65 cm) and eluted with water. The fractions containing **1** were pooled and lyophilized from water to afford **1** (300 mg, 58% in 2 steps) as a colorless amorphous powder (HCl salt): ^1H and ^{13}C NMR spectra were in good agreement with those reported;^{1b,2a} ^1H NMR (D_2O) δ 1.91–2.02 (m, 1H, H-5), 2.88 (t, 1H, $J = 11.9$ Hz), 2.97 (t, 1H, $J = 12.7$ Hz), 3.48–3.56 (m, 3H), 3.71–3.85 (m, 3H); ^{13}C NMR (D_2O) δ 43.6, 45.6, 49.4, 61.8, 71.1, 73.9; HRMS FAB calcd for $\text{C}_6\text{H}_{13}\text{NO}_3$ ($M + \text{H}^+$) 148.0974, found 148.0976.

***tert*-Butyl (3R,4R,5S)-5-[(*tert*-Butyldiphenylsilyloxy)]-3,4-dihydroxypiperidine-1-carboxylate (11).** A mixture of **9** (1.6 g, 2.9 mmol) and Raney Ni (3 mL) in EtOH (20 mL) was stirred vigorously under H_2 atmosphere at room temperature

for 50 h. After removal of the catalyst by filtration, the filtrate was concentrated to give a crude product **10**, which was employed for the next step without further purification.

A mixture of the residue **10**, Boc₂O (1.2 g, 5.8 mmol), and Et₃N (0.8 mL, 5.8 mmol) in MeOH (30 mL) was stirred for 12 h at room temperature and concentrated. The residue was chromatographed on silica gel with EtOAc–hexane (1:4) to give **11** (850 mg, 62% in 2 steps) as a colorless oil: ¹H NMR (CDCl₃) δ 1.07 (9H, s), 1.45 (9H, s), 1.79 (br s, 1H), 2.53 (m, 2H), 3.49 (br s, 2H), 3.78 (br s, 2H), 4.03–4.16 (m, 1H), 4.25 (br s, 1H), 7.40–7.44 (m, 6H), 7.65–7.68 (m, 4H); ¹³C NMR (CDCl₃) 19.1, 26.8, 28.3, 46.5, 71.6, 80.1, 127.8, 129.9, 135.5 (d), 154.4; HRMS FAB calcd for C₂₇H₃₉NO₅Si (M + H)⁺ 486.2676, found 486.2684.

tert-Butyl (3*R*,4*R*,5*S*)-5-(Hydroxymethyl)-3,4-di-*O*-methoxymethyl-3,4-dihydropiperidine-1-carboxylate (12). To a stirred solution of **11** (0.5 g, 1.0 mmol) and *i*Pr₂NEt (0.5 mL, 2.8 mmol) in CH₂Cl₂ (15 mL) was added MOMCl (0.23 mL, 3.0 mmol) at 0–5 °C, and the mixture was allowed to warm to room temperature and stirred for 30 h at room temperature. The reaction mixture was poured into ice-cooled aqueous NaHCO₃ solution (150 mL) and extracted with Et₂O. The combined extracts were successively washed with aqueous NaHCO₃ solution and brine, dried (MgSO₄), and concentrated. The residue was employed for the next step without further purification.

A mixture of the above residue and Bu₄NF (1 M solution in THF, 1.3 mL, 1.5 mmol) in THF (20 mL) was stirred for 4 h at room temperature. The reaction mixture was poured into H₂O (30 mL)–EtOAc (50 mL), and the aqueous layer was re-extracted with EtOAc. The combined extracts were successively washed with aqueous NaHCO₃ solution and brine, dried (MgSO₄), and concentrated. The residue was chromatographed on silica gel with EtOAc–hexane (1:2) to give **12** (340 mg, 74% in two steps) as a colorless oil: ¹H NMR (CDCl₃) δ 1.38 (s, 9H), 1.63 (br s, 1H), 2.91–2.94 (br s, 2H), 3.31 (s, 3H), 3.36 (s, 3H), 3.45–3.48 (m, 3H), 3.78–3.86 (m, 2H), 4.62 (m, 3H), 4.82 (d, 1H, *J* = 6.6 Hz); ¹³C NMR (CDCl₃) δ 28.2, 42.9, 55.4, 56.0, 60.4, 79.9, 95.9, 98.3, 154.7; HRMS FAB calcd for C₁₅H₃₀NO₇ (M + H)⁺ 336.2022, found 336.2024.

tert-Butyl (3*R*,4*R*,5*S*)-5-Carboxyl-3,4-di-*O*-methoxymethyl-3,4-dihydropiperidine-1-carboxylate (13). A solution of DMSO (0.17 mL, 2.4 mmol) in CH₂Cl₂ (5 mL) was added dropwise to a cooled solution of (COCl)₂ (0.14 mL, 1.6 mmol) in CH₂Cl₂ (20 mL) at –78 °C, and the mixture was stirred for 30 min. To this mixture was added dropwise a solution of **12** (270 mg, 0.8 mmol) in CH₂Cl₂ (10 mL) at –78 °C, and the resulting mixture was stirred another 1.5 h at the same temperature. A solution of Et₃N (0.6 mL, 4.0 mmol) in CH₂Cl₂ (10 mL) was then added to the reaction mixture, which was gradually warmed to 0–5 °C over 30 min. The reaction mixture was poured into ice water and extracted with EtOAc. The combined extracts were successively washed with water, aqueous NaHCO₃ solution, and brine, dried (MgSO₄), and concentrated to give an aldehyde derivative, which was employed for the next step without further purification.

To a mixture of the above aldehyde and NaH₂PO₄·H₂O (1.1 g, 8.0 mmol) in CH₃CN (15 mL)–H₂O (3 mL) were successively added a 35% H₂O₂ solution (0.09 mL, 1.0 mmol) and a solution of NaClO₂ (80%, 110 mg, 1.0 mmol) in water (10 mL) at 0–5 °C, and the resulting mixture was stirred for 1 h at room temperature. The reaction was terminated by the addition of Na₂SO₃ (100 mg), and the resulting mixture was extracted with EtOAc. The combined extracts were successively washed with water and brine, dried (MgSO₄), and concentrated. The residue was chromatographed on silica gel with CHCl₃–MeOH (50:1) to give **13** (230 mg, 82% in two steps) as a colorless oil: ¹H NMR (CDCl₃) δ 1.40 (s, 9H), 2.58 (ddd, 1H, *J* = 4.2, 9.3, 12.9 Hz), 2.85 (dd, 1H, *J* = 9.3, 13.2 Hz), 3.10 (br s, 1H), 3.30 (s, 3H), 3.33 (s, 3H), 3.48 (br s, 1H), 3.86 (dd, 1H, *J* = 8.4 Hz), 3.98–4.08 (m, 2H), 4.62–4.67 (m, 3H), 4.80 (d, 1H, *J* = 6.0 Hz); ¹³C NMR (CDCl₃) δ 28.1, 47.3, 55.5, 55.9, 77.2, 78.1, 80.5, 96.1, 97.4, 154.4, 175.2; HRMS FAB calcd for C₁₅H₂₈NO₈ (M + H)⁺ 350.1815, found 350.1815.

D-Glucuronic Acid Type 1-*N*-Iminosugar: (3*R*,4*R*,5*S*)-5-Carboxylpiperidine-3,4-diol Hydrochloride Salt (2). A solution of **13** (100 mg, 0.29 mmol) in 3 N HCl (15 mL) was stirred overnight at room temperature and concentrated. The residue was chromatographed on silica gel with 2-propanol–H₂O–NH₄OH (7:2:1) to give chromatographically pure **2**. Water (10 mL) and 1 N HCl (20 mL) were added to the residue, and the solution was concentrated to form a hydrochloride salt of the iminosugar **2**. The iminosugar **2** was further purified by a column of Sephadex G-25 (2 cm × 65 cm) eluted with water. The fractions containing **2** were pooled and lyophilized from water to afford **2** (50 mg, 89%) as a colorless amorphous powder (HCl salt): NMR data of **2** were in good accordance to those reported;^{1b} ¹H NMR (D₂O) δ 2.77 (ddd, 1H, *J* = 4.2, 7.4, 7.8 Hz, H-5), 2.94 (dd, 1H, *J* = 7.5, 12.9 Hz, H-2ax), 3.24 (dd, 1H, *J* = 7.8, 13.2 Hz, H-6ax), 3.34 (dd, 1H, *J* = 3.9, 12.9 Hz, H-2eq), 3.38 (dd, 1H, *J* = 5.1, 13.2 Hz, H-6eq), 3.76 (ddd, 1H, *J* = 3.3, 7.2, 7.7 Hz, H-3), 3.93 (t, 1H, *J* = 6.9 Hz, H-4); ¹³C NMR (D₂O) δ 44.2, 46.2, 47.9, 69.0, 71.9, 175.7; HRMS FAB calcd for C₆H₁₂NO₄ (M + H)⁺ 162.0766, found 162.0766.

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Supporting Information Available: ¹H and ¹³C NMR spectra for compounds **1–6**, **8**, **9**, and **11–13**. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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